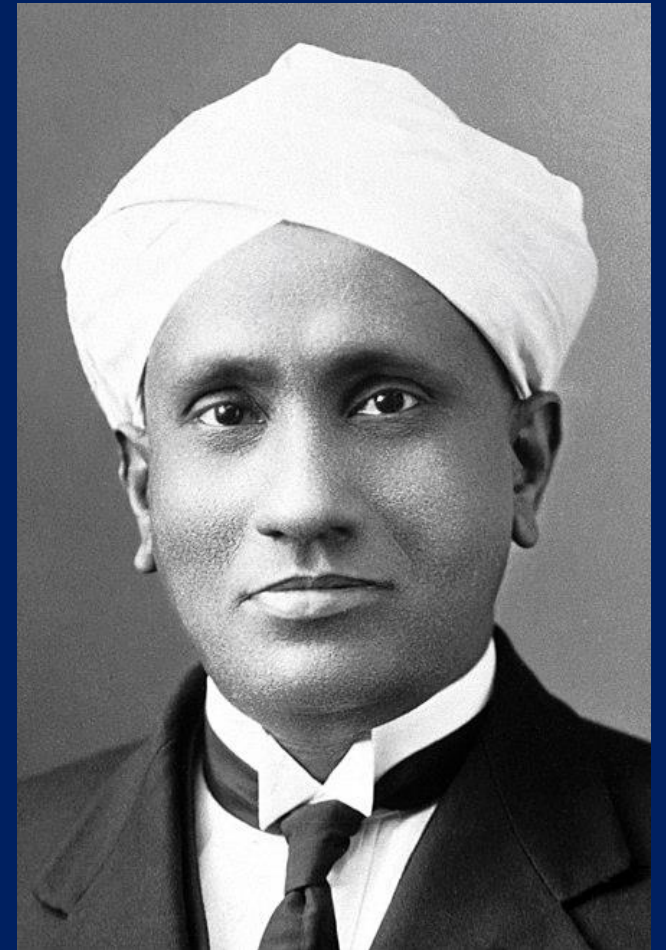
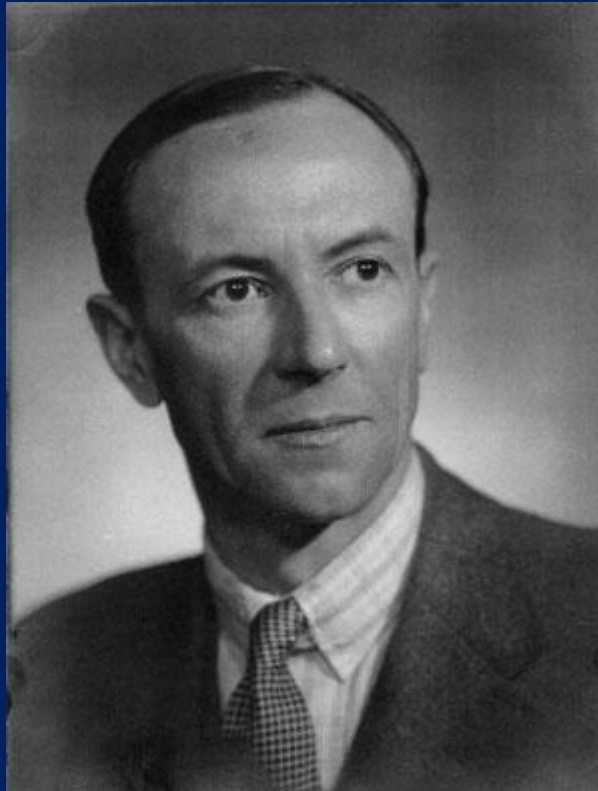


CropBooster

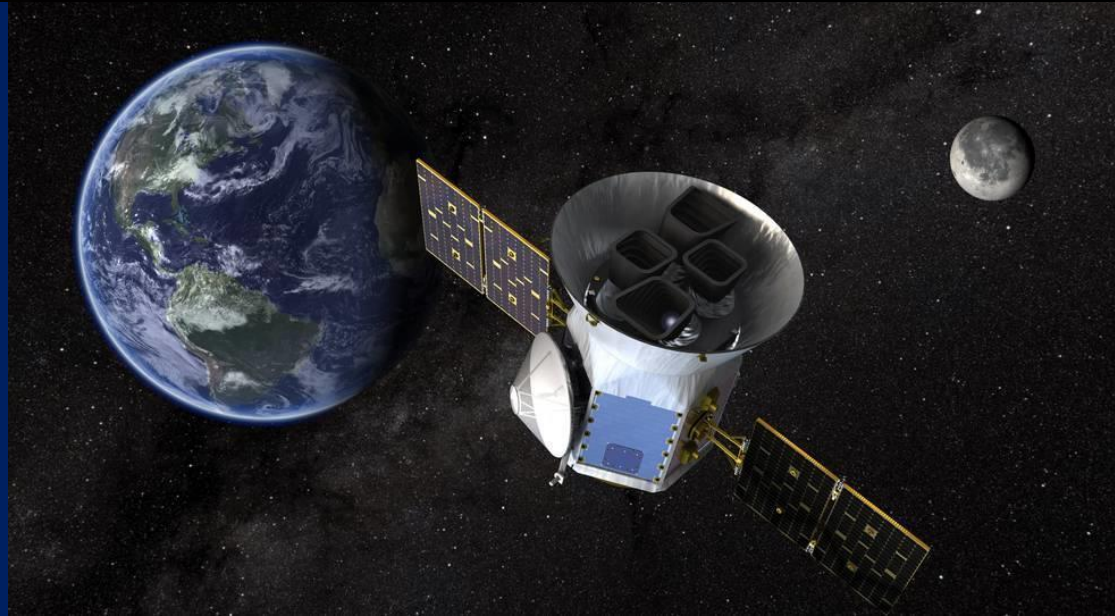
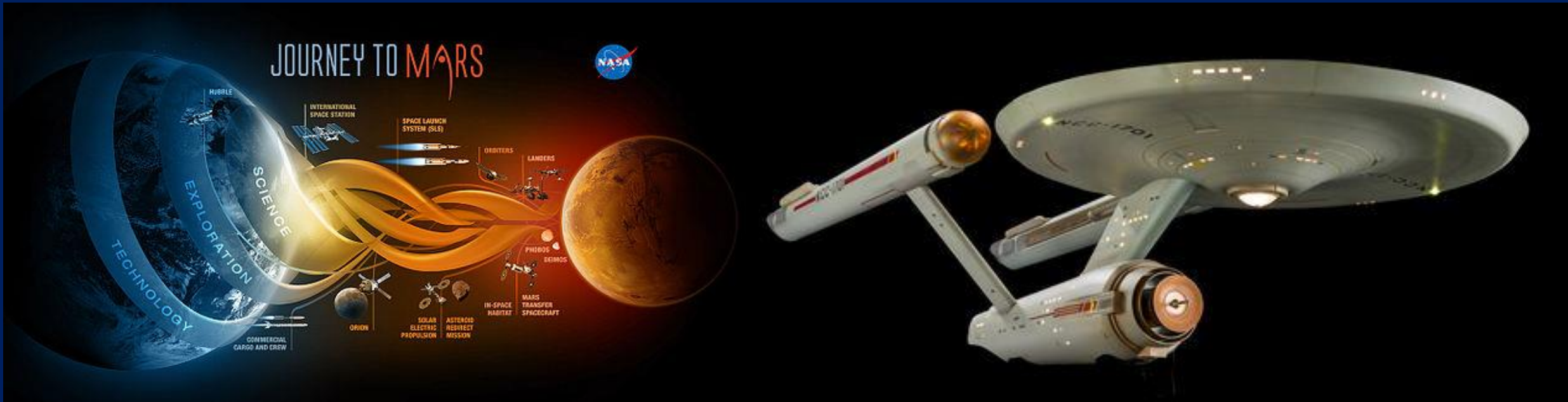
Our road to the future of phenomics: The actors are Neutron, Raman and X-Ray



CropBooster

Our road to the future of phenomics: The actors are Neutron, Raman and X-Ray





State of the Art Technologies for Plant Phenotyping



Research
&
Development



Sensors

3D Imaging

Kinetic chlorophyll
fluorescence Imaging

Hyperspectral Imaging

X-RAY

Raman Spectroscopy

Neutron Imaging

Mass Spectroscopy

Root Imaging

RGB and Morphometric
Imaging

Near Infrared/Thermal
Imaging

LIBSpectroscopy



Sensors

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Kinetic chlorophyll
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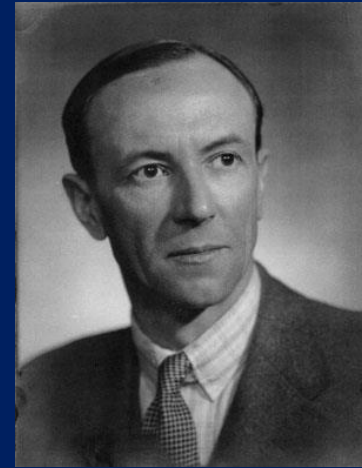
Near Infrared/Thermal
Imaging

LIBSpectroscopy

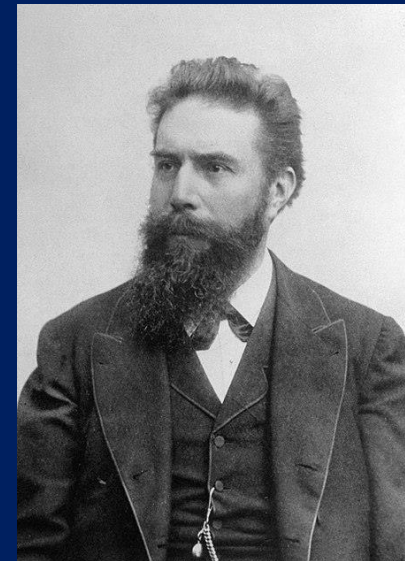


Martin Trtilek

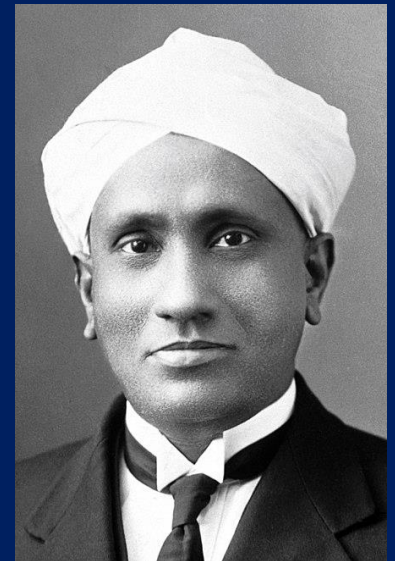
Photon Systems Instruments



Chadwick



Roentgen



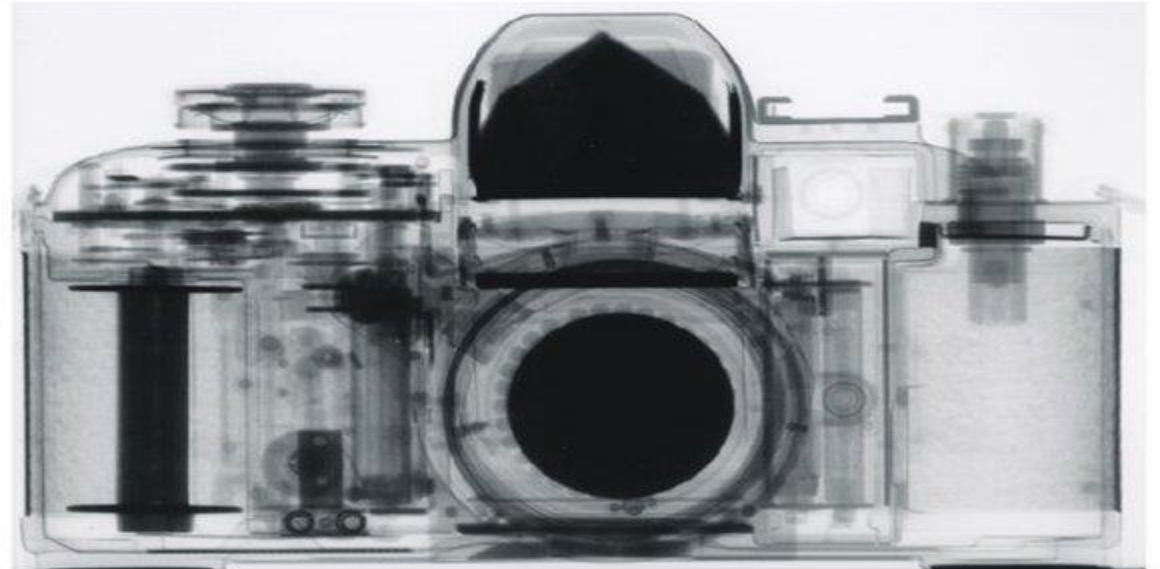
Raman

Look inside

Comparison of X-ray and Neutron Radiographs



X-ray



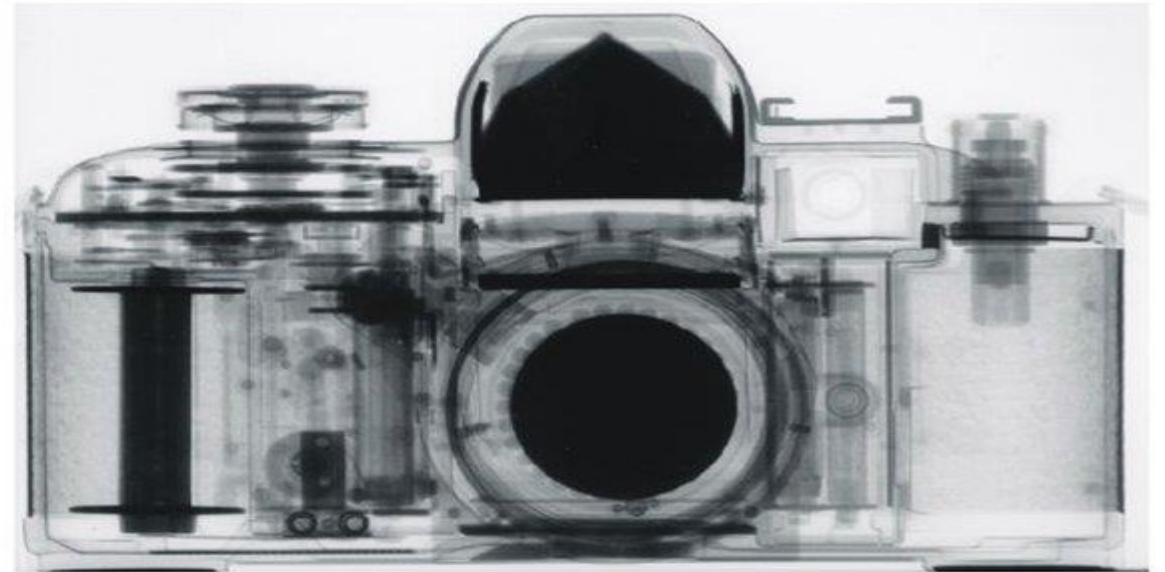
Neutrons

X-rays interact with an atom's electron cloud, while neutrons interact with the nucleus—which means that they interact differently with different materials. Neutrons will pass through some materials that X-rays cannot whereas X-rays will more easily pass through materials that neutrons cannot.

Comparison of X-ray and Neutron Radiographs

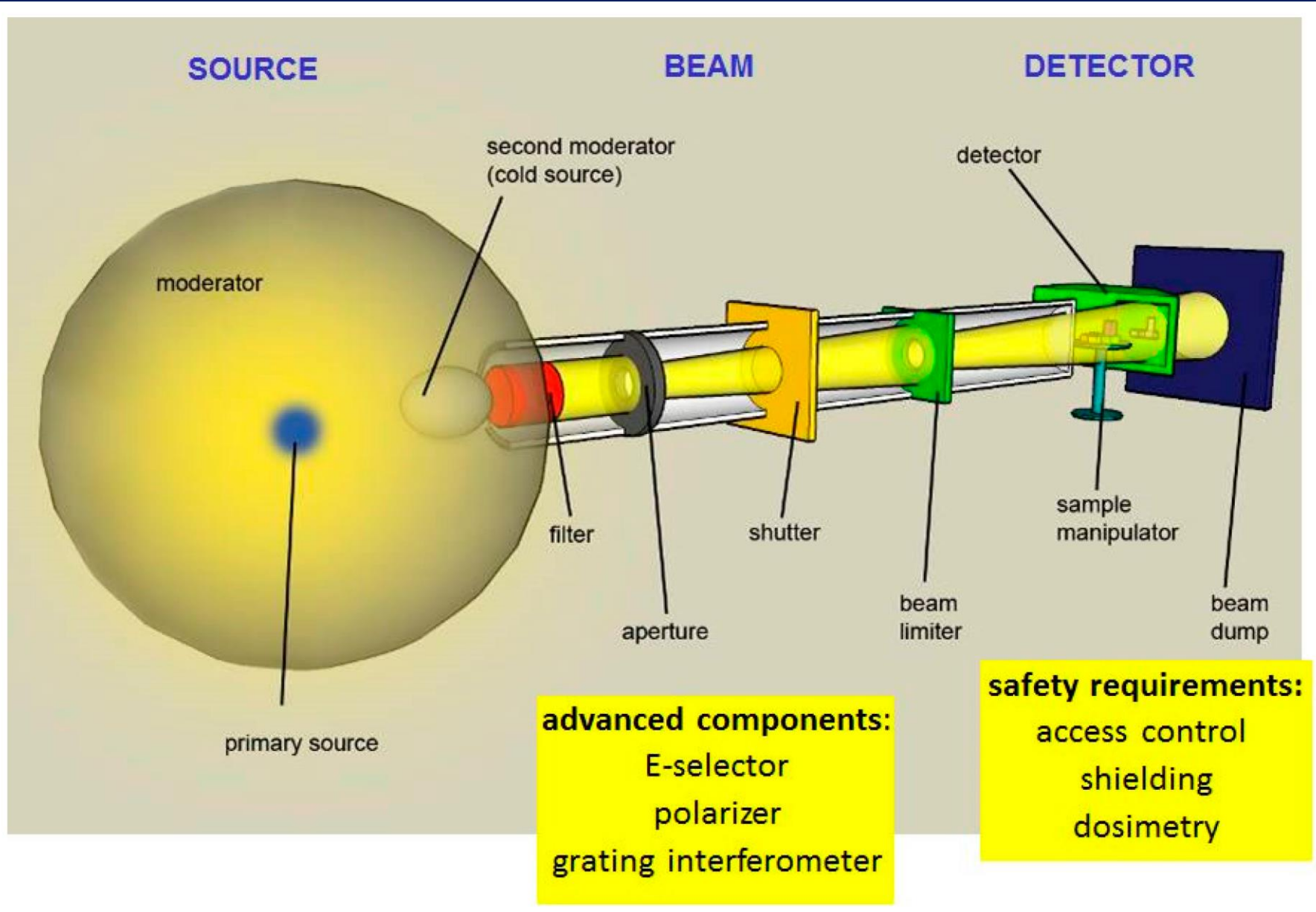


X-ray



Neutrons

Neutron imaging



Neutron imaging

- Neutron imaging is a non-invasive non-destructive imaging technique
- Neutron imaging can provide two- or three-dimensional, spatially resolved images of the internal structure of bulk samples that are not accessible by other techniques
- Due to the interaction properties of neutrons with matter, some light nuclei such as H and D greatly **scatter neutrons**, whereas some heavier elements such as Al, Si (e.g., SiO₂ – sand), Cu or Pb are not strong scatterers or absorbers of neutrons and can therefore be easily **penetrated**.¹
- Neutron radiography measures the attenuation of a neutron beam caused by absorption and scattering within a sample using a two-dimensional position-sensitive detector that measures the transmitted neutron flux.¹

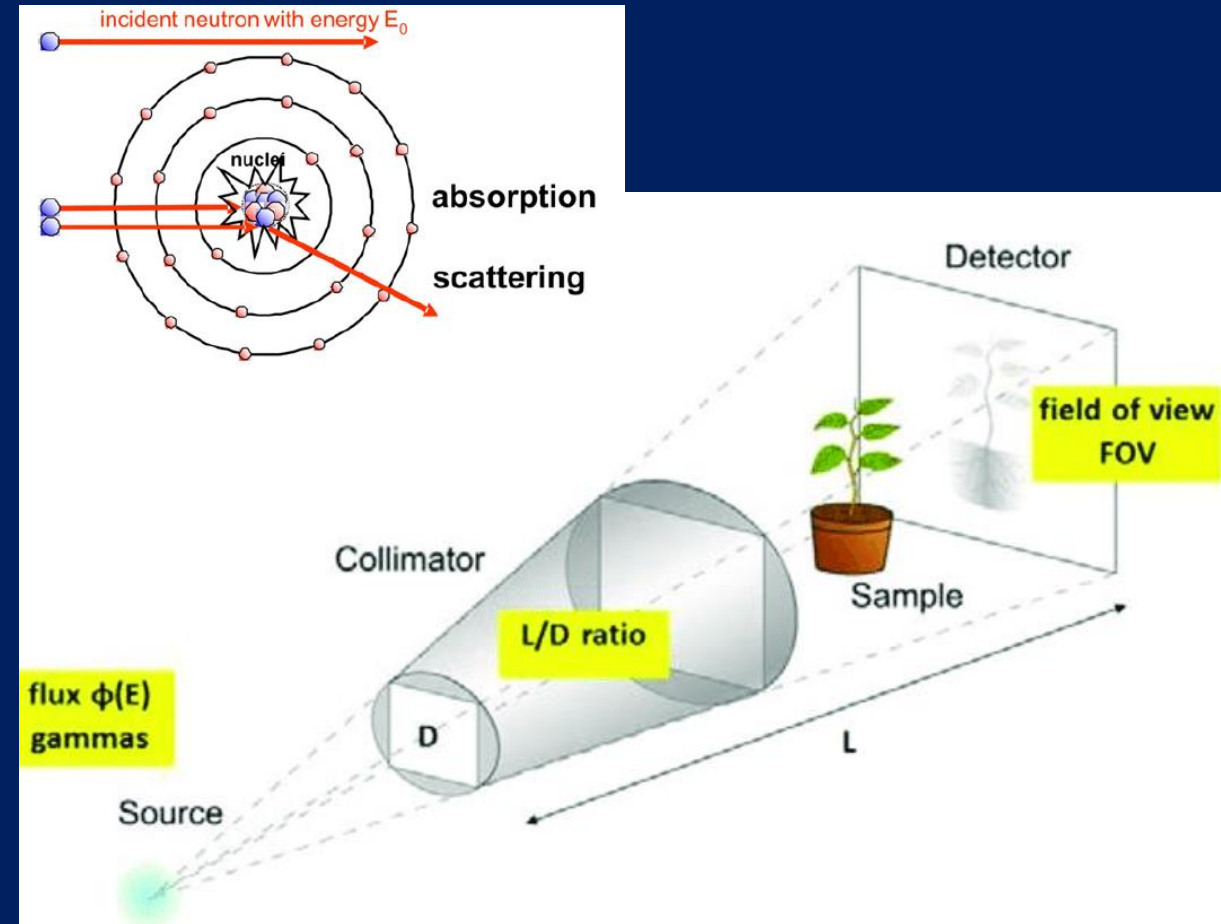


Figure: Neutron Imaging in the radiography mode: principle setting and essential parameters for the image quality determination²

1. M Strobl et al 2009 J. Phys. D: Appl. Phys. 42 243001

2. Lehmann, Eberhard & Raventos, Marc & Harti, Ralph & Trtik, Pavel & Kaestner, Anders & Mannes, David & Grünzweig, C.. (2017). Methodical Progress in Neutron Imaging at PSI. Physics Procedia. 88. 250-257. 10.1016/j.phpro.2017.06.035.

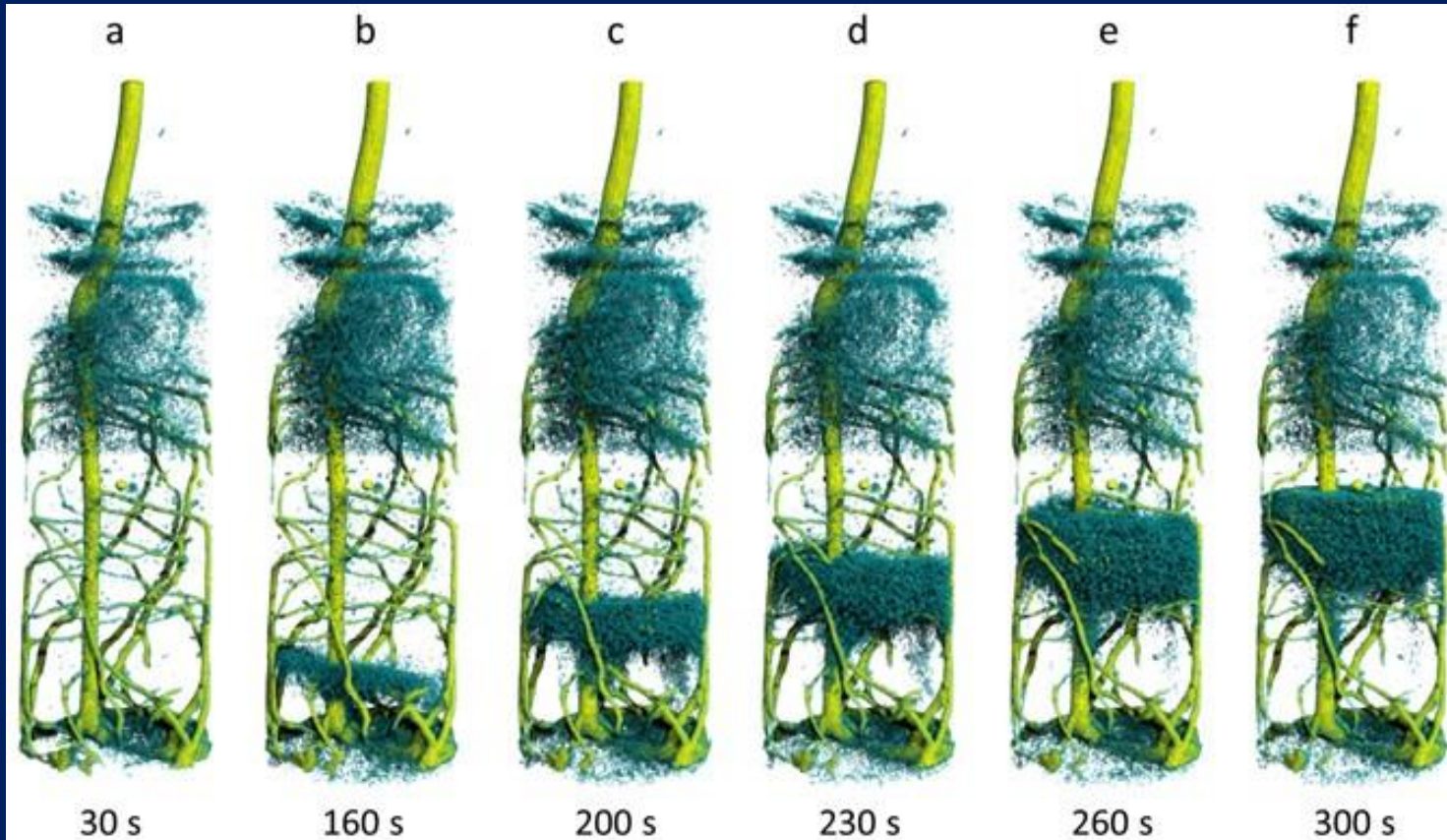


Figure: Time-resolved neutron tomography of the lupine root system after the injection of 4 ml deuterated water (D₂O) through the bottom.

The time series ($30 \text{ s} \leq t \leq 300 \text{ s}$) shows the ascending front of water (H₂O) displaced by deuterated water moving upwards. The time resolution for each tomogram is just 10 s.

Capturing 3D Water Flow in Rooted Soil by Ultra- fast Neutron Tomography

Christian Tötze, Nikolay Kardjilov, Ingo Manke & Sascha E. Oswald

Neutron radiography and computed tomography of biological materials with complex geometries

BER STRUCTURAL BIOLOGY AND IMAGING RESOURCES

Synchrotron, Neutron, and Cryo-EM

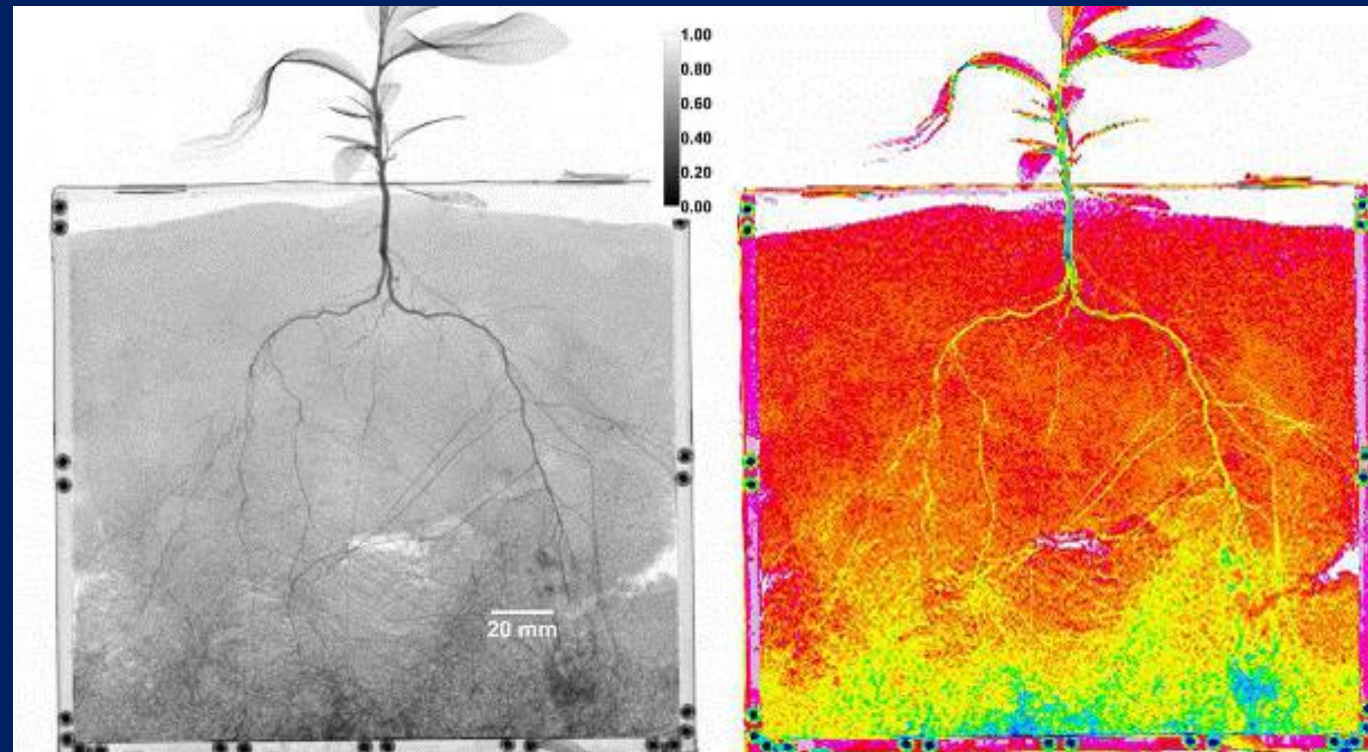
U.S. Department of Energy | Office of Science | Office of Biological and Environmental Research

Key Features of Neutron Imaging

- Spatial resolution of 10 to 50 μm
- High-penetrating and non-destructive
- Hydrogen/deuterium isotope contrast
- Accommodates versatile sample types including organisms, soil, rock, and experimental devices
- Boron, lithium, or gadolinium contrast agents

BER Researchers Use Neutron Imaging to Study:

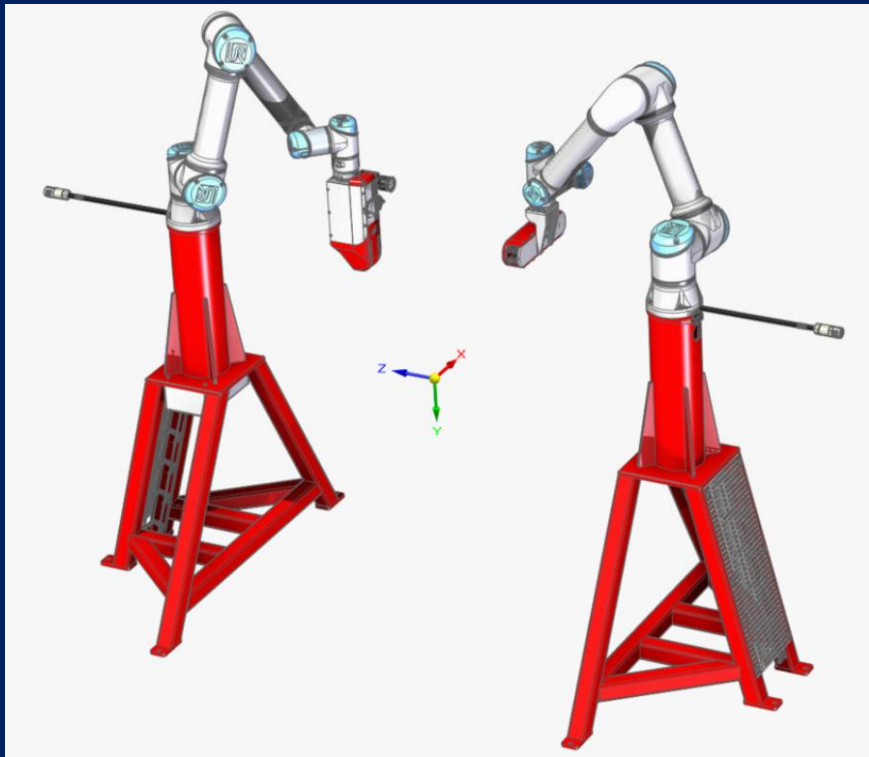
- Water uptake by root systems
- Water transport in soils
- Soil structure
- Rhizosphere water dynamics
- Adaptation of root systems in the presence of pollutants
- Root behavior in extreme environments (e.g., drought)
- Belowground root system response to climate change



Water uptake under stress. Neutron radiography of a black cottonwood (*Populus trichocarpa*) planted in sand enabled examination of differences in water uptake by old and new plant roots following recovery from extreme drought stress. [Springer Nature - Dhiman I., et al. 2017. "Quantifying root water extraction after drought recovery using sub-mm *in situ* empirical data," *Plant and Soil* 424, 1–17. DOI: 10.1007/s11104-017-3408-5. Copyright 2017.]

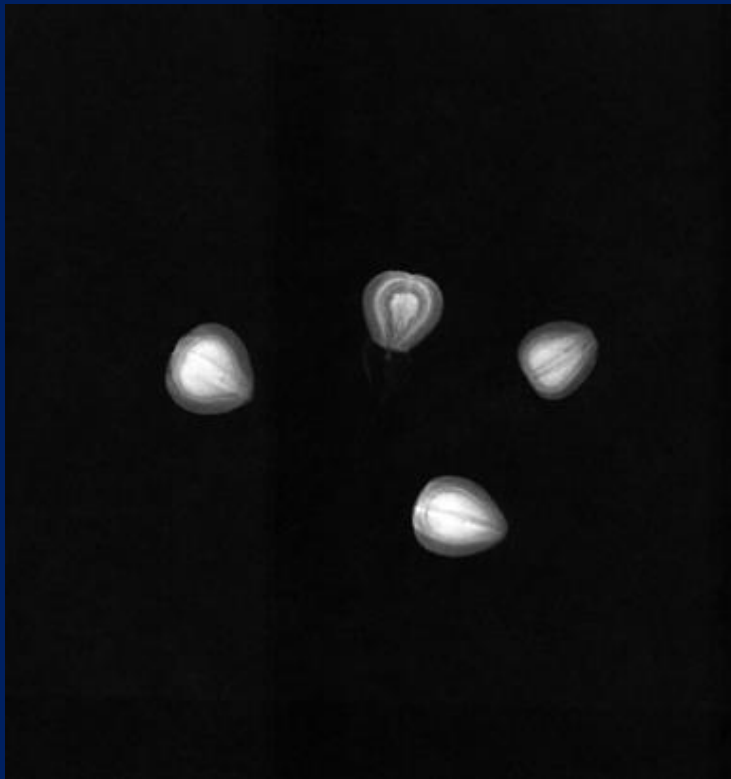
X-ray imaging

- X-ray imaging system combines single particle counting X-ray imaging detectors for high quality images with the flexibility of collaborative robots.
- The key parts of the scanner are two robotic arms with 6 joints. The first arm holds an X-ray tube, the second holds an imaging detector.
- The major benefit of the system is the viewing angle flexibility, superior image contrast and resolution.

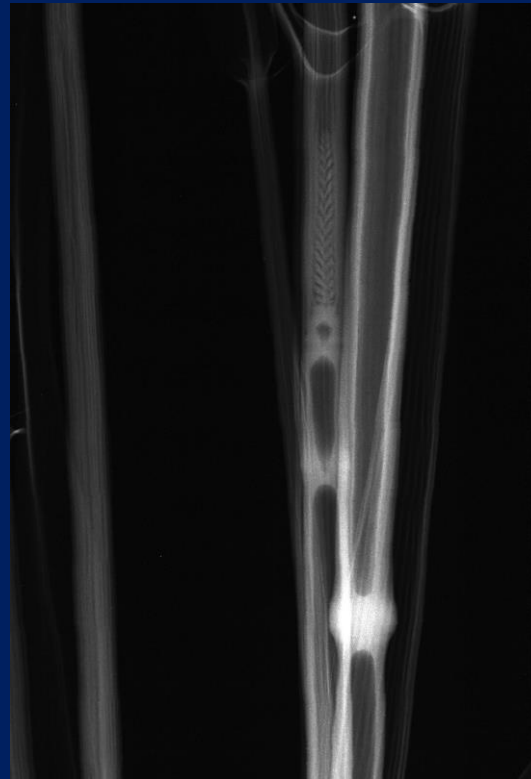


X-ray imaging application

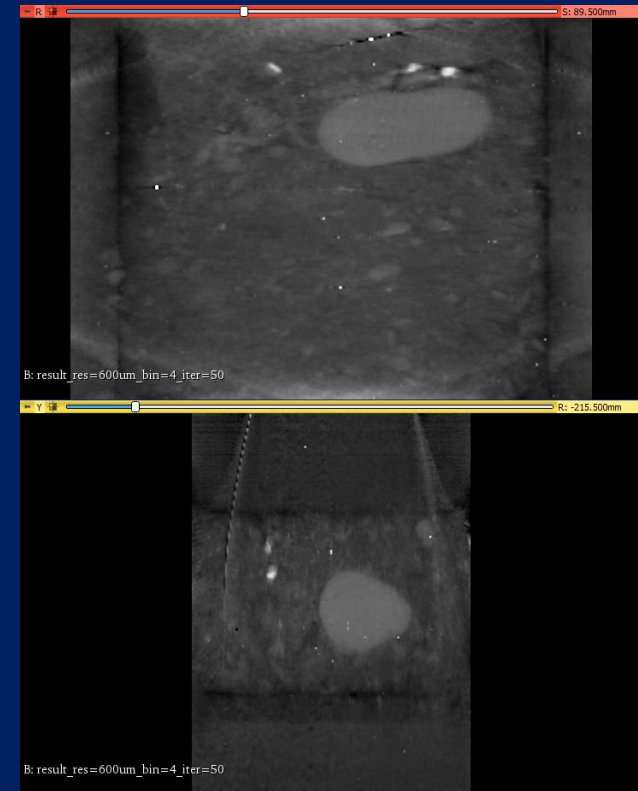
- Uncover inner structures, areas concealed by the leaf canopy, exact biomass of the stalks and ears,
- X-ray computed tomography (CT) used in the analysis of underground structures - to analyse and interpret subtle volumetric changes of belowground organs such as tubers



Seed quality analysis

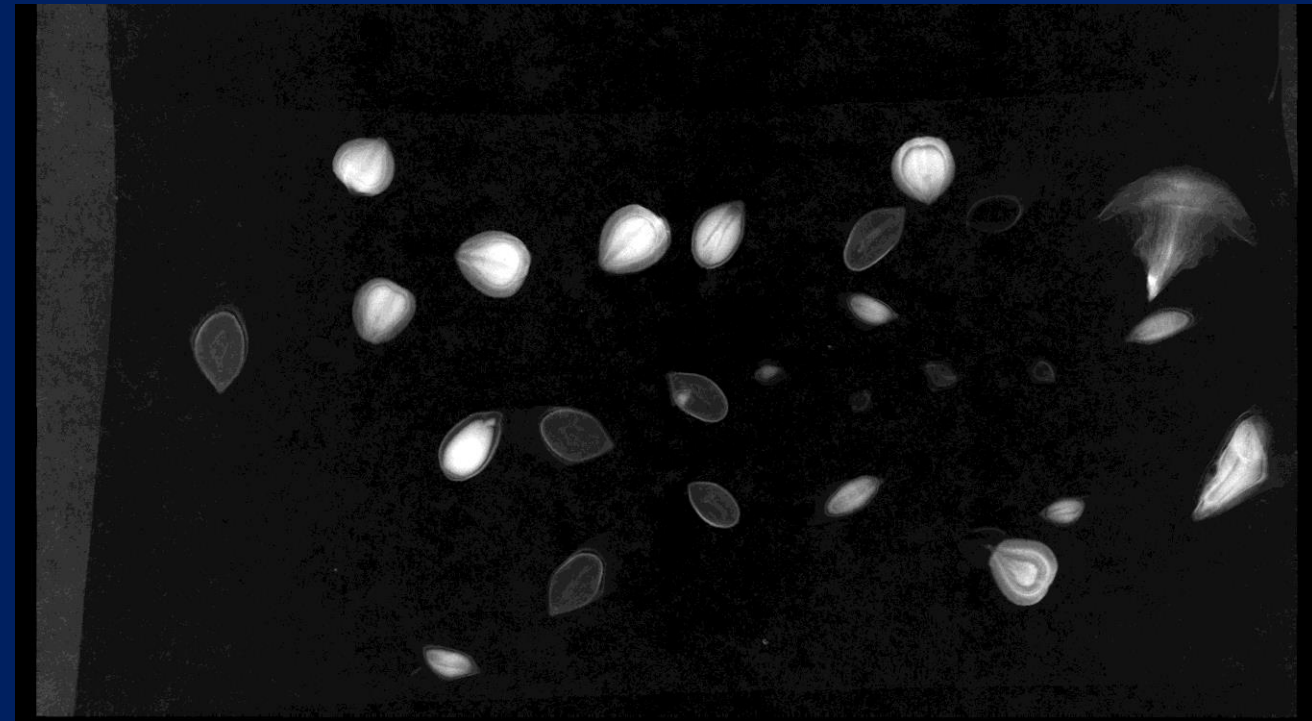


Spike development analysis



Tuber and root structure analysis

High-scale seed quality analysis



Non-destructive spikelet development and grain filling analysis

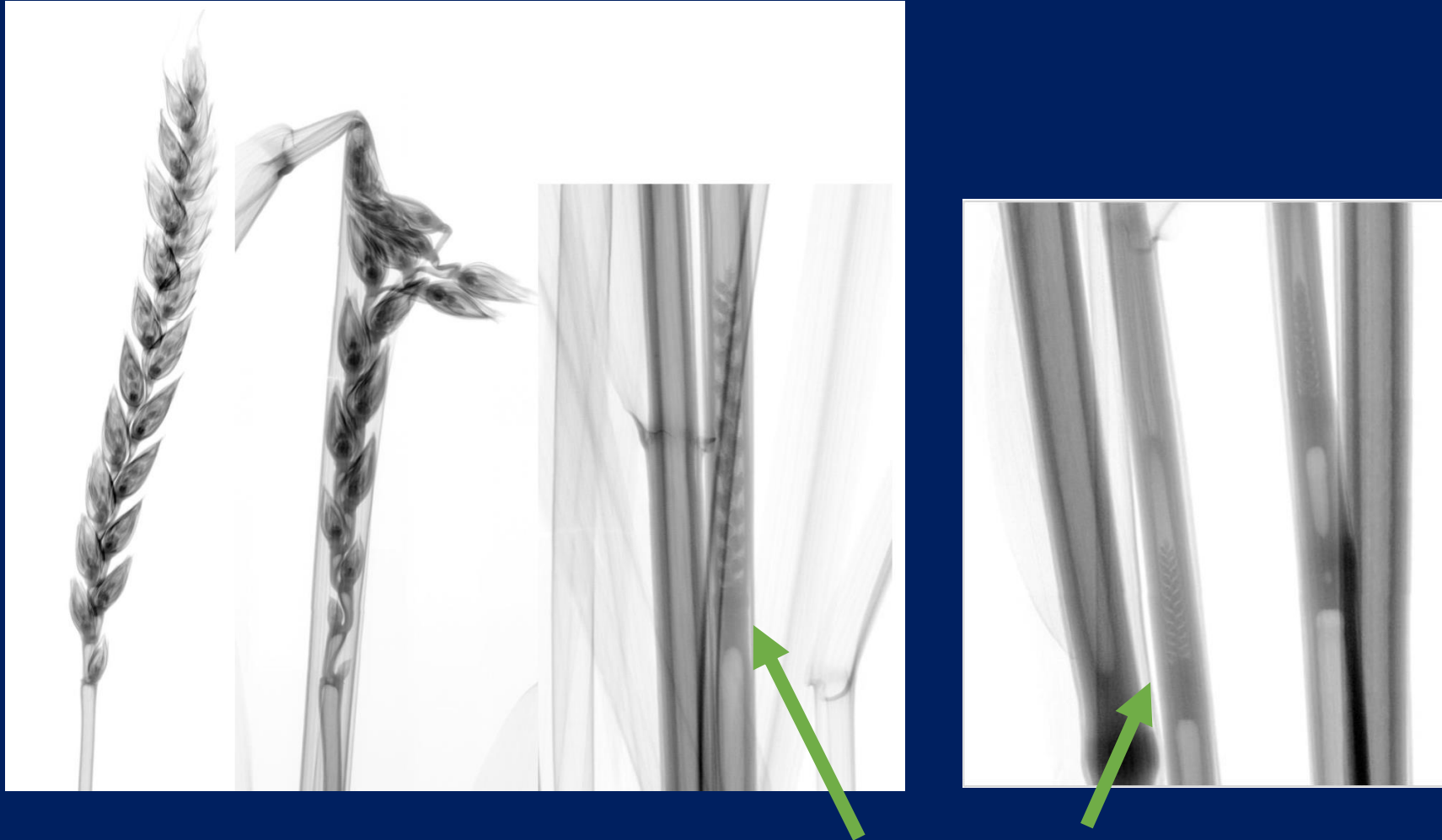


RGB image showing two ears without obvious defects

X-ray image shows that some grains in the spike are empty

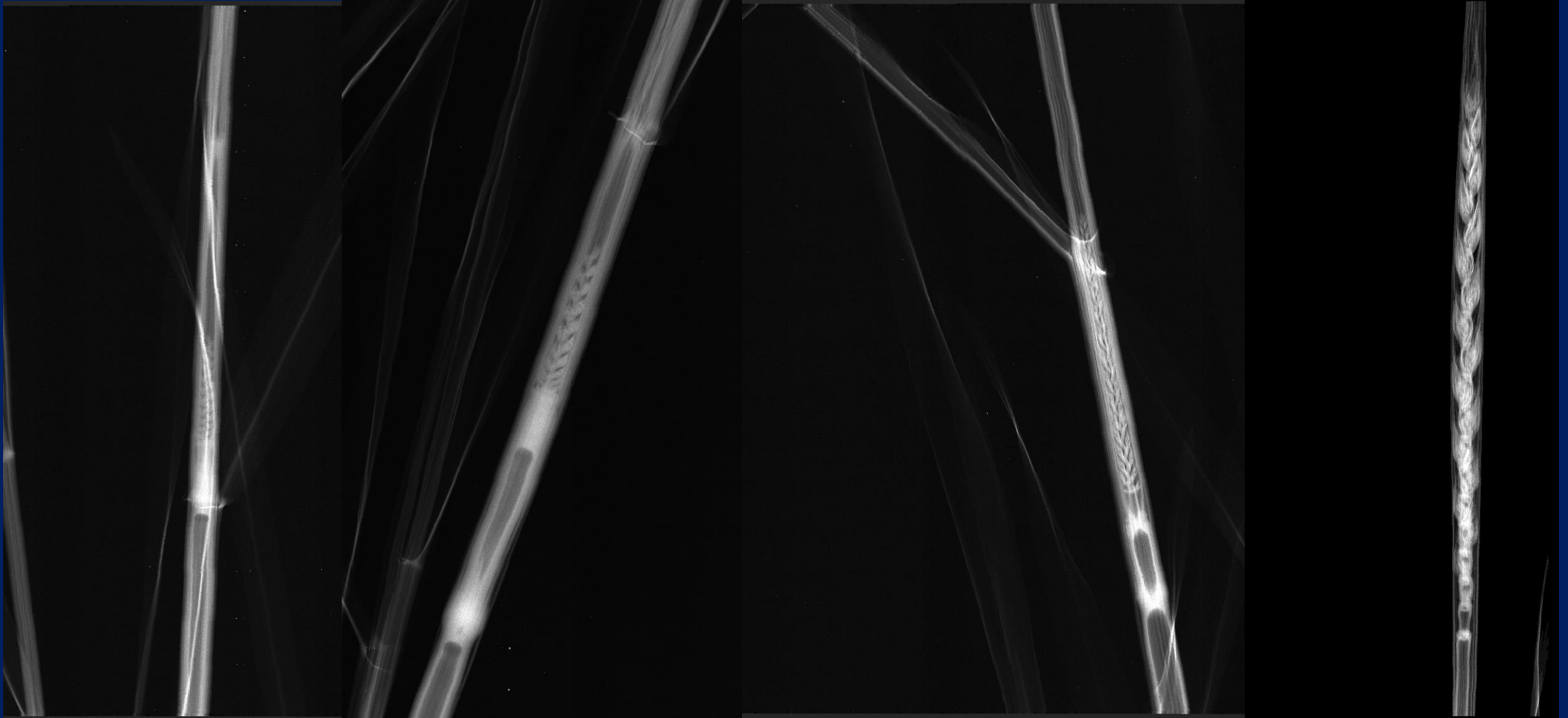


Non-destructive image-based characterisation of apical meristem and spike developmental stages

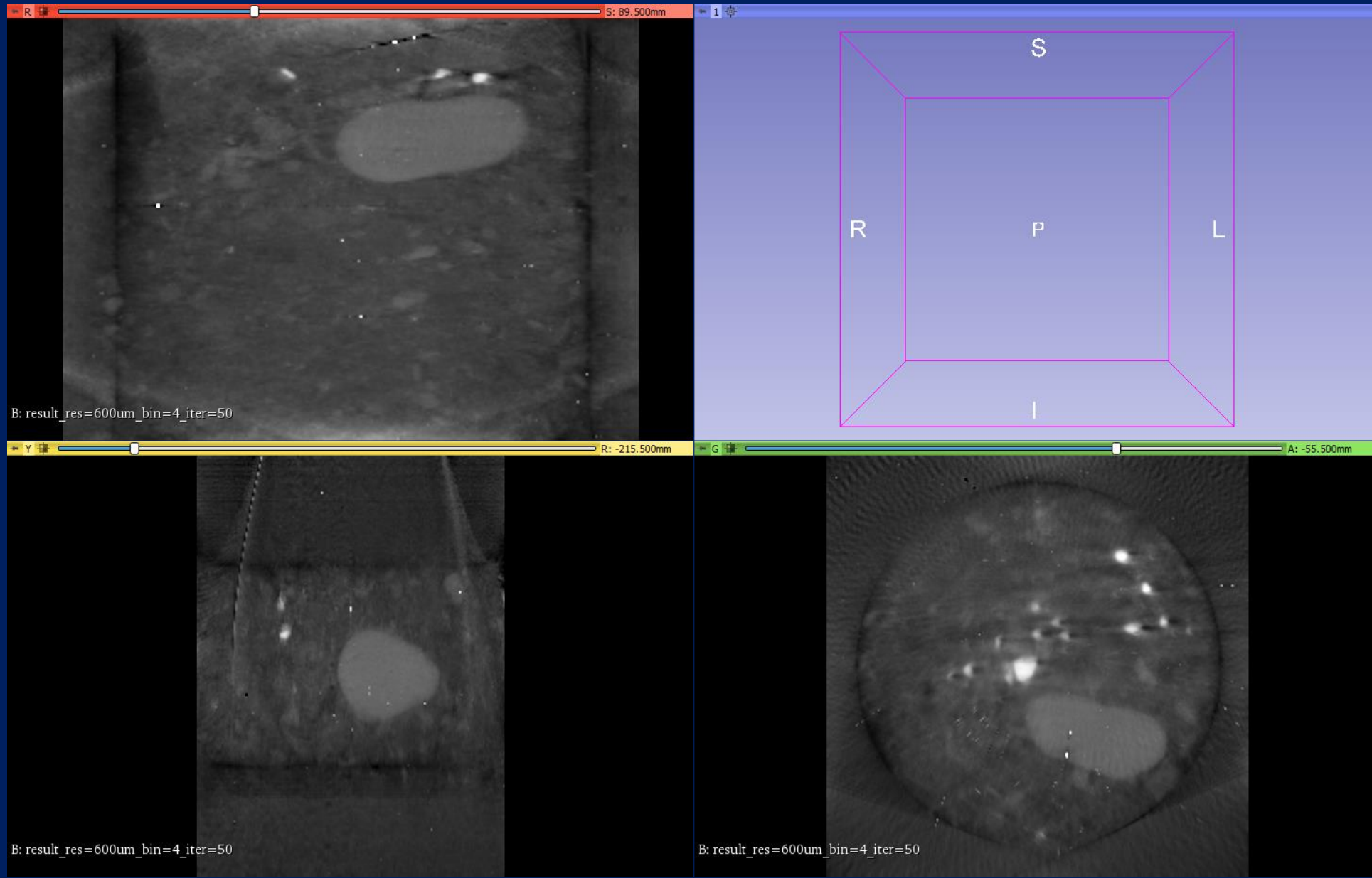


Early stages of spikelet development

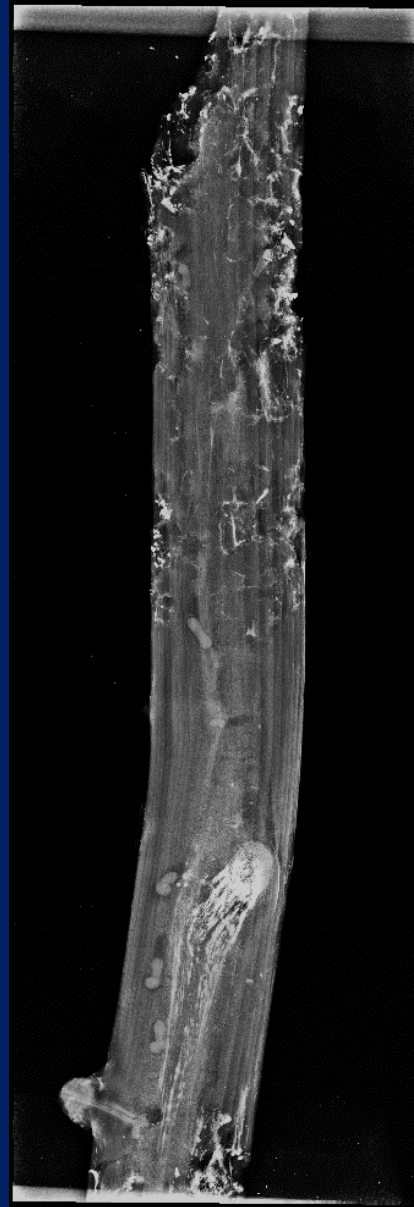
Non-destructive image-based characterisation of apical meristem and spike developmental stages



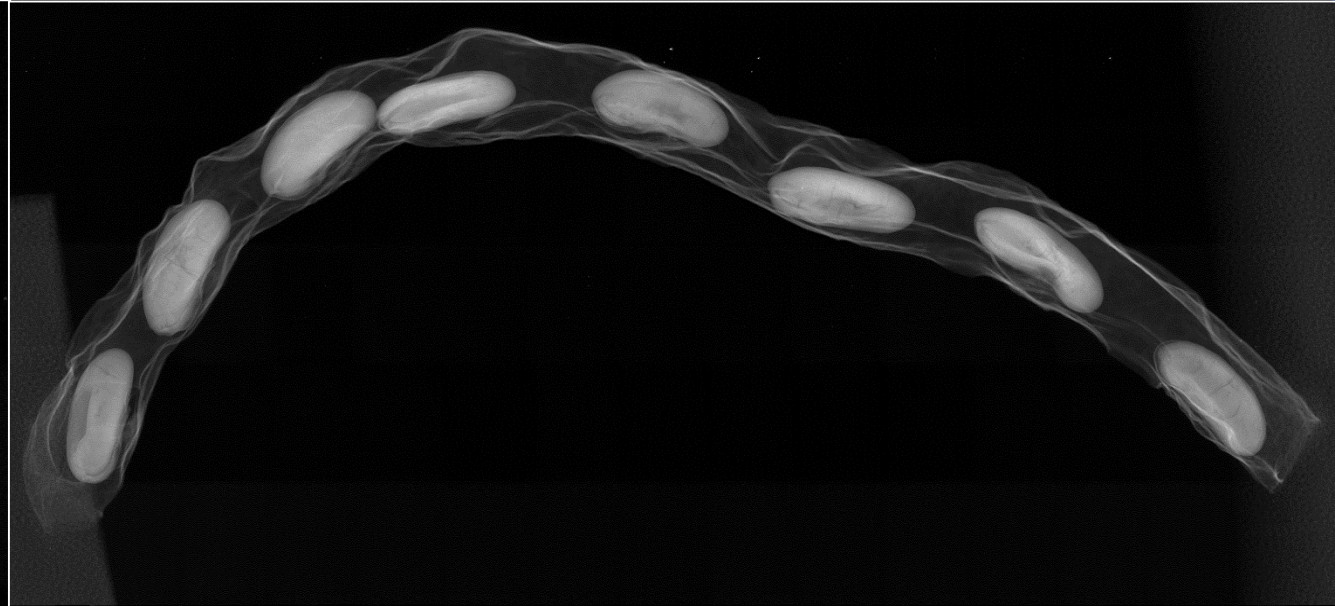
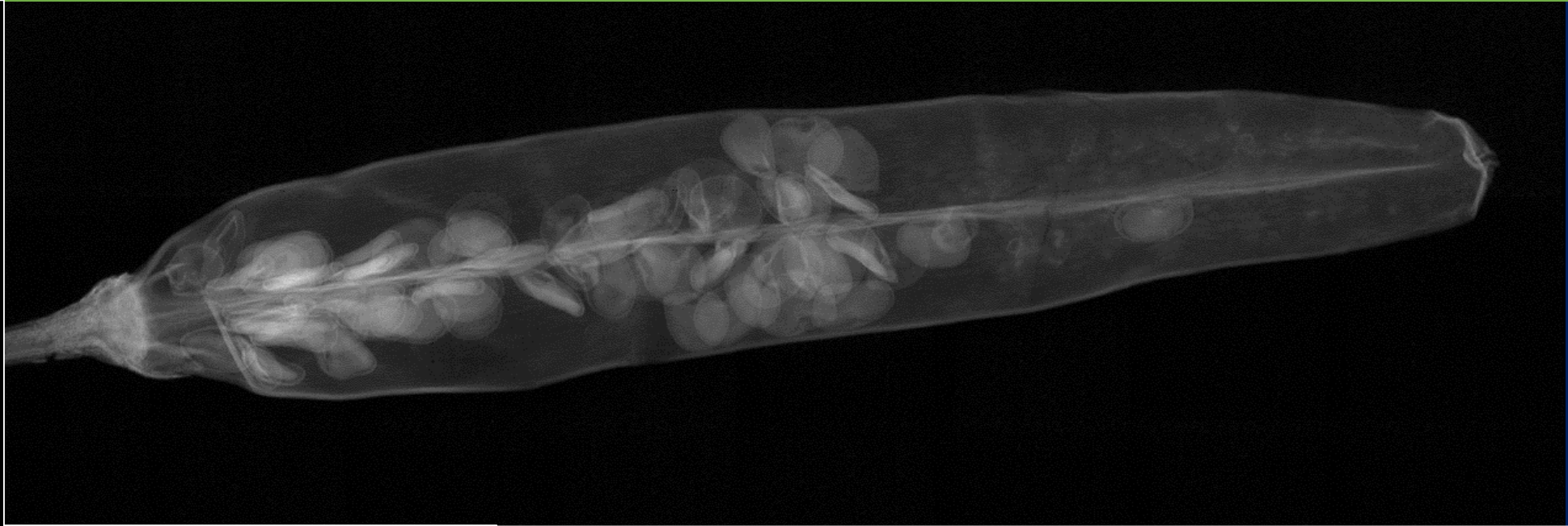
Tuber and root structure analysis

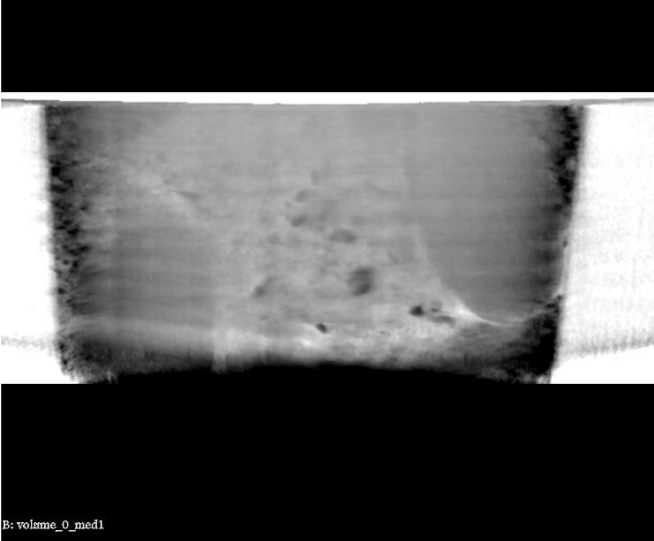
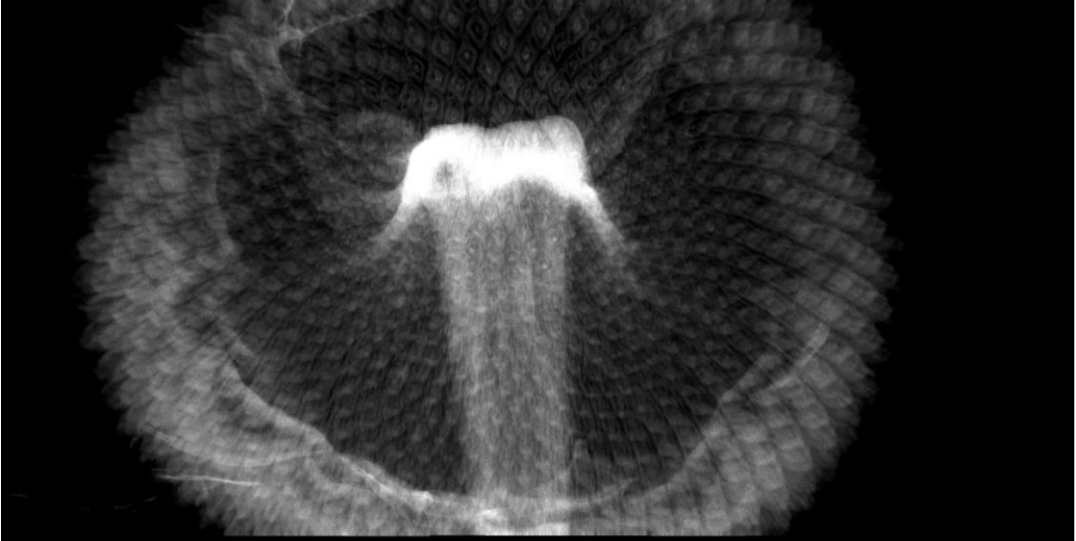
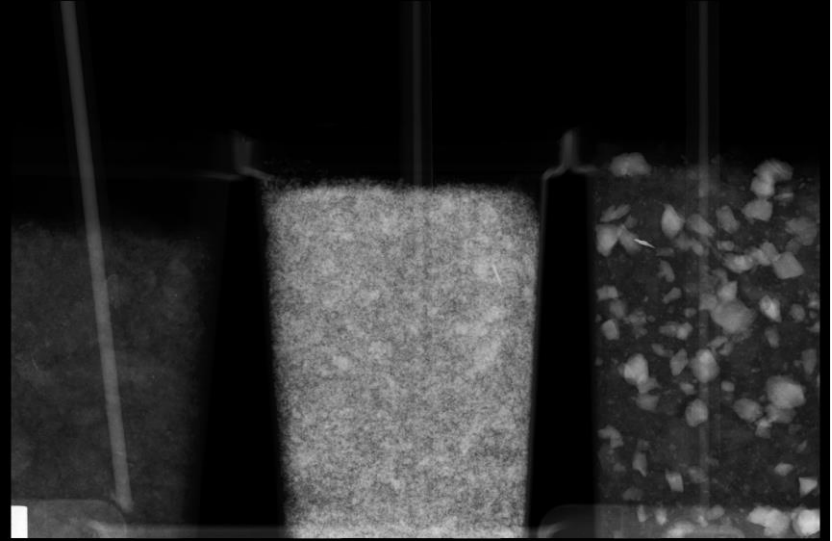
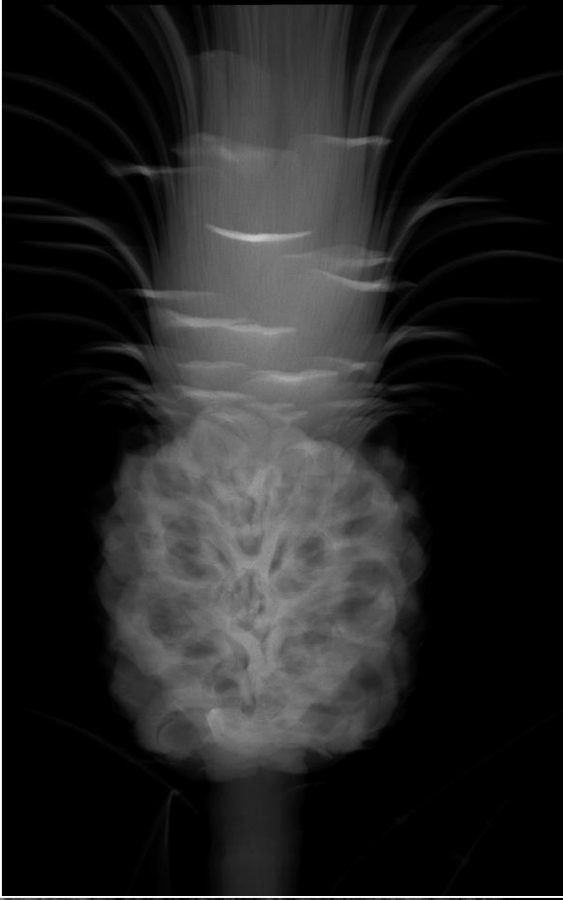


Inner structure of the objects

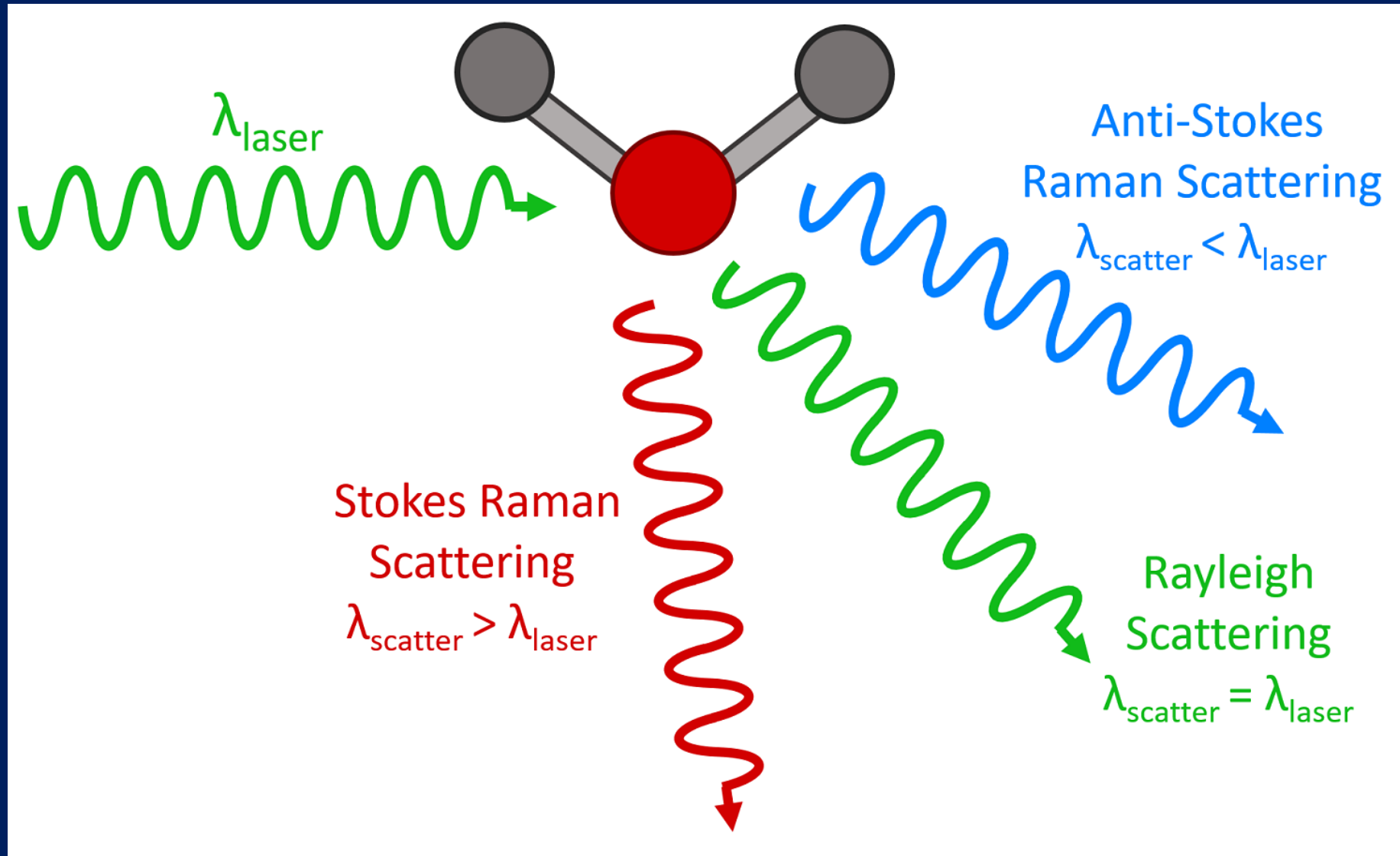


X-RAY - Morphometric Imaging

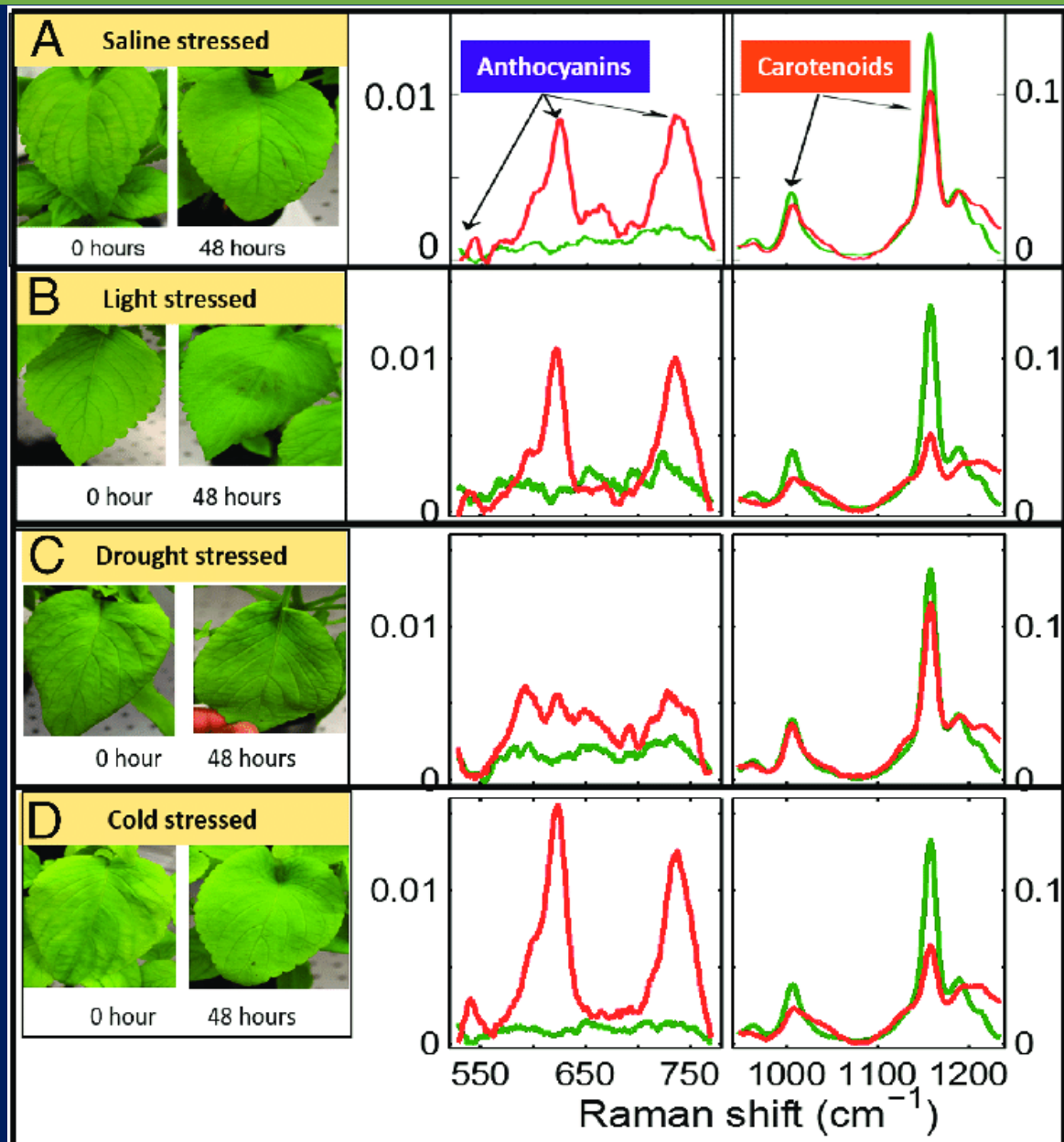




Raman - spectroscopy



Raman - spectroscopy

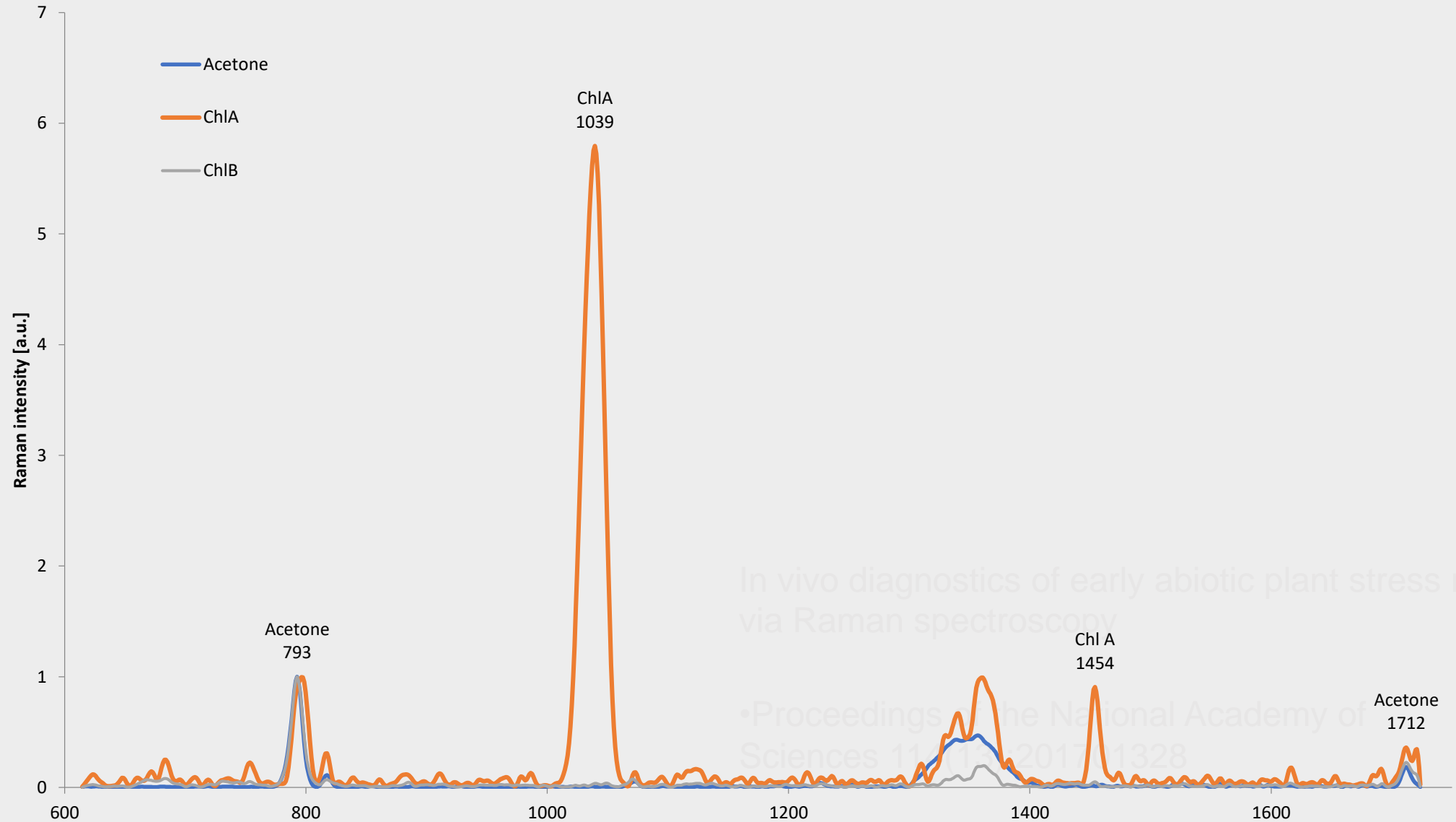


Molecular fingerprint

In vivo diagnostics of early abiotic plant stress response via Raman spectroscopy

Proceedings of the National Academy of Sciences 114(13):201701328

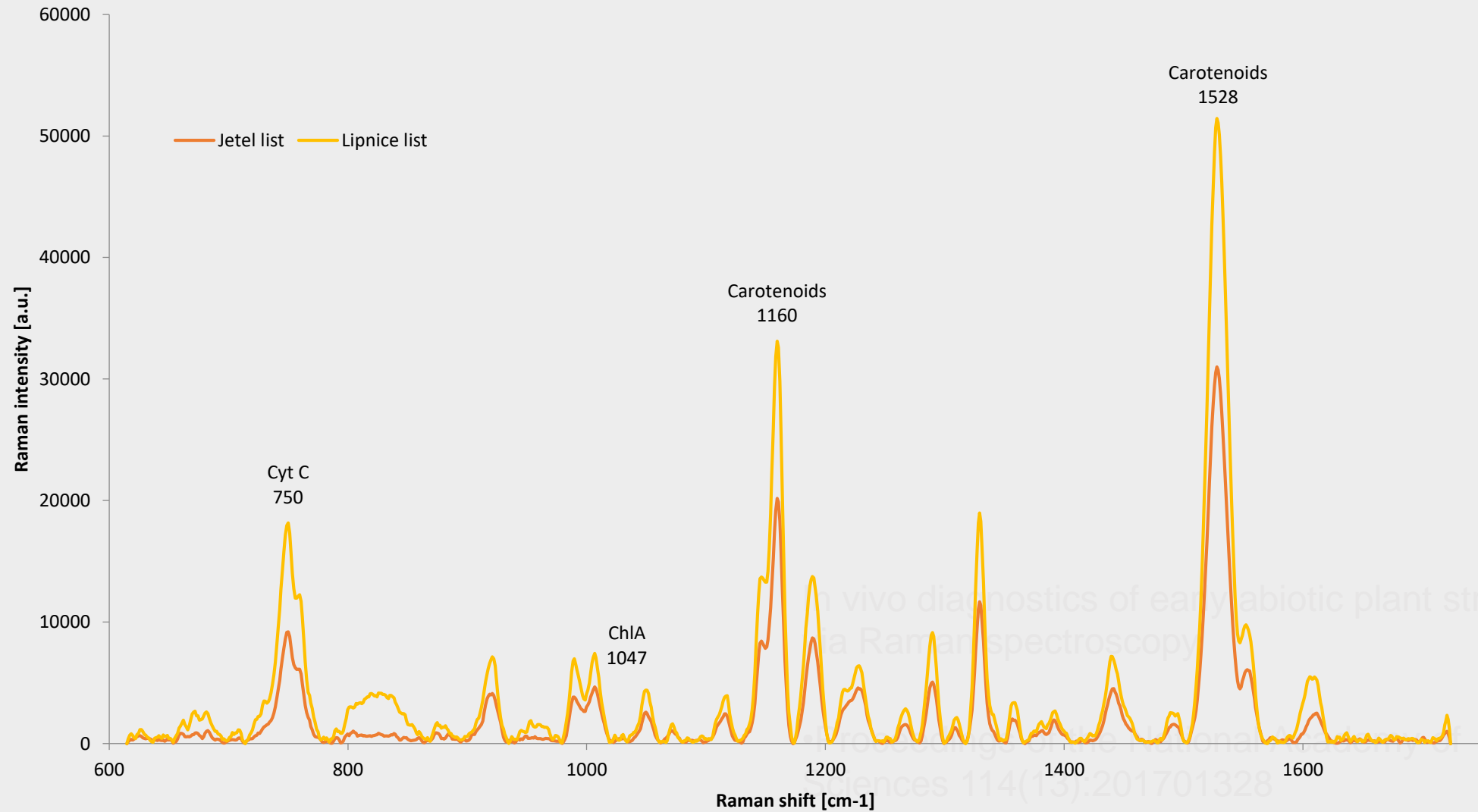
Raman – Chlorophyll A and B



In vivo diagnostics of early abiotic plant stress response
via Raman spectroscopy

• Proceedings of the National Academy of
Sciences 114 (2017) 1328

Raman – Chla – carotenoids – real samples



In vivo diagnostics of early abiotic plant stress response via Raman spectroscopy
Proceedings of the National Academy of Sciences 114(13):201701328

Object of interest
recognition
from
RGB
Using AI



Leading of Raman
detector to the
measuring spot